



ABSTRACT

Itch is an unpleasant sensation that emanates primarily from the skin. The chemical mediators that drive neuronal activity originate from a complex interaction between keratinocytes, inflammatory cells, nerve endings, and the skin microbiota, relaying itch signals to the brain. Stress also exacerbates itch via the skin-brain axis. Recently, the microbiota has surfaced as a major player to regulate this axis, notably during stress settings aroused by actual or perceived homeostatic challenge. The routes of communication between the microbiota and brain are slowly being unraveled and involve neurochemicals (i.e., acetylcholine, histamine, catecholamines, and corticotropin) that originate from the microbiota itself. By focusing on itch biology and by referring to the more established field of pain research, this review examines the possible means by which the skin microbiota contributes to itch.

KEYWORDS: Itch, skin microbiota, stress, microbiota-skin-brain axis

THE SKIN MICROBIOTA AND ITCH: Is There a Link?

by HEI SUNG KIM and GIL YOSIPOVITCH

Dr. Kim is with the Department of Dermatology and Cutaneous Surgery at Miami Itch Center, Miller School of Medicine at University of Miami in Miami, Florida, the Department of Dermatology at Incheon St. Mary's Hospital, The Catholic University of Korea in Seoul, Korea, and the Department of Biomedicine and Health Sciences, at The Catholic University of Korea in Seoul, Korea. Dr. Yosipovitch is with the Department of Dermatology and Cutaneous Surgery at Miami Itch Center, Miller School of Medicine at the University of Miami in Miami, Florida.

Originally published in *J Clin Med*. 2020 Apr 22;9(4):1190. doi: 10.3390/jcm9041190. Reprinted and distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>). Minor revisions to the text were made to adhere to journal style.

Bacteria, viruses, fungi, archaea, helminths, and protozoa that inhabit our body are a prospering dynamic community shaping a symbiotic superorganism. Roughly 1,014 microbiota populate the entire body, with their number approximating that of human cells.^{1,2} Evidence suggests that microbiota take part in maintaining human health.^{3,4}

As a crucial barrier to the exterior world, skin is the body's largest organs.⁵ A square centimeter of human skin holds around 10^6 of microbiota.⁶⁻⁸ The symbionts defend against illness by regulating the skin barrier and host immune response.^{9,10} On the other hand, microbial imbalance (dysbiosis) has been noted to exacerbate skin lesions and delay wound healing.^{11,12} Recently, the emerging role of the skin microbiota in itch has received attention.¹³ Large-scale changes of the skin microbiota have been noted in itchy skin diseases. *Staphylococcus aureus* (*S. aureus*) participates in atopic dermatitis (AD) flare-up; its colonization correlates with disease severity and itch.¹⁴⁻¹⁶

In the present review, we offer an integrative perspective on the skin microbiota and itch. The first section describes the interplay of the cutaneous microbiota with the epidermal barrier, the local immune system, and the sensory nerve, proposing the microbiota's peripheral mechanism of itch. The second section concentrates on the concept of microbial endocrinology and addresses the microbiota-skin-brain axis. Moreover, the interaction between the skin microbiota and the

amygdala is discussed to explain the microbiota's central mechanism of itch. Overall, this article describes the putative role of the skin microbiota in itch.

THE PERIPHERAL MECHANISM LINKING THE SKIN MICROBIOTA AND ITCH

Itch arises from the activation of epidermal nerve fibers that belong to a specialized class of itch-provoking neurons ("pruriceptors"). The chemical mediators that drive neuronal activity arise from complex interaction between keratinocytes, inflammatory cells, and nerve endings, coupled with upregulated immune cascades, epidermal barrier dysfunction, and exogenous environmental stimuli (e.g., microbiota, allergens, irritants).¹⁷ Peripheral nerves relay cues from the skin to the dorsal root and trigeminal ganglia and then to the spinal cord and brain where central itch processing takes place (Figure 1).¹⁷

Skin barrier. The skin barrier shields the body from a wide range of external dangers.¹⁸ It consists of the epidermis and several layers below that harbor microbiota.¹⁹⁻²¹ The physical skin barrier is the stratum corneum, which comprises dead keratinocytes and proteinaceous crosslinking filaments.²²

The skin also has a chemical barrier of antimicrobial peptides (AMPs) that are constitutively expressed or induced. AMPs directly block microbial growth or provoke the immune

FUNDING: This study was supported by a National Research Foundation of Korea (NRF) grant founded by the South Korean Government (2017R1C1B5016144).

DISCLOSURES: Yosipovitch reports serving on the scientific board and being a consultant for Trevi, Sanofi Regeneron, Galderma, Pfizer, Novartis, Kiniksa, Eli Lilly, Bellus, LEO and is supported by Sun Pharma, Pfizer, Novartis, Kiniksa, Leo Pierre Fabre. None of these involvements had influence on the content of the presented paper.

CORRESPONDENCE: Gil Yosipovitch; Email: gyosipovitch@med.miami.edu

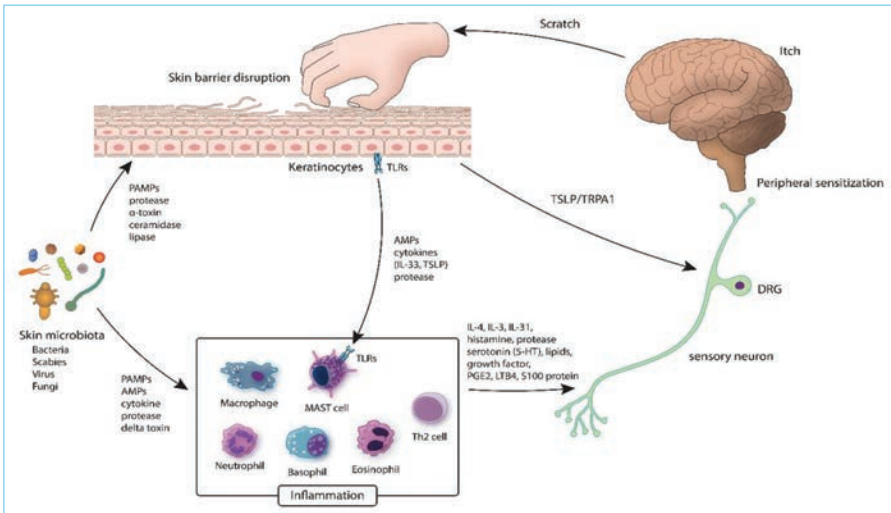


FIGURE 1. Inflammatory circuit of the skin microbiota. Various microbiota (bacteria, fungi and viruses) cover the exterior of a healthy skin where the barrier is intact. In the event of dysbiosis, pathogens release proteases, which may disrupt the epidermal barrier. Delta-toxin causes mast cell degranulation, which prompt inflammation and itching. AMP: antimicrobial peptides; DRG: dorsal root ganglia; IL: interleukin; LTB4: leukotriene B4; PAMP: pathogen associated molecular pattern; PGE2: prostaglandin E2; TLR: Toll-like receptor; TRPA1: transient receptor potential antigen 1; TSLP: thymic stromal lymphopoietin. Figure 1. Inflammatory circuit of the skin microbiota. Various microbiota (bacteria, fungi and viruses) cover the exterior of a healthy skin where the barrier is intact. In the event of dysbiosis, pathogens release proteases, which may disrupt the epidermal barrier. Delta-toxin causes mast cell degranulation, which prompt inflammation and itching. AMP: antimicrobial peptides; DRG: dorsal root ganglia; IL: interleukin; LTB4: leukotriene B4; PAMP: pathogen associated molecular pattern; PGE2: prostaglandin E2; TLR: Toll-like receptor; TRPA1: transient receptor potential antigen 1; TSLP: thymic stromal lymphopoietin.

reaction. One example is the liberation of histamine and prostaglandin D₂ (PG-D₂)²³ by mast cells in respect to human β -defensins (h β Ds) and LL-37, causing pruritus.

The skin microbiota is an integral part of the skin barrier.¹⁸ It protects the host from pathogens by competing for nutrients and space.¹⁹ Some produce antimicrobial compounds that block the growth of competitors.¹⁹ Symbionts also alter the skin barrier via bacterial enzymes, such as proteases, which impact corneocyte desquamation, and lipases, which break down skin surface lipids.²⁴

Staphylococcus epidermidis (*S. epidermidis*) is the primary bacterium colonizing the human epithelia and is a vital member of the skin resident microbiota.²⁵ *S. epidermidis* has a flexible interrelation with its host and deposits biofilms (a physical barrier) that are remarkably hard to clear.²⁶ Symbiont strains of *S. epidermidis* suppress *S. aureus* biofilm formation by producing serine protease (Esp), which also enhances the antimicrobial effect of h β Ds.²⁷ Another typical skin resident is *Cutibacterium acnes* (*C. acnes*), which inhibits the growth of methicillin-resistant *S. aureus* (MRSA).²⁸ In short, *C. acnes* ferments glycerol, a natural metabolite in human skin, into

short-chain fatty acids (SCFAs) that maintain an acidic skin pH.²⁹ Symbionts flourish at acidic pH, whereas potential pathogens, such as *S. aureus*, thrive at neutral pH.^{30,31}

Intrinsic (host) and extrinsic (environmental) factors affect skin barrier function by shaping microbial structure.³² *S. aureus* colonization is found in up to 90 percent of patients with AD.³³ It produces ceramidase, which breaks down ceramides, an essential component of the skin barrier.³⁴ *S. aureus* also produces toxins that impede wound healing and bring epithelial barrier disintegration.³⁵ Scabies mites (*Sarcoptes scabiei*) alter the skin microbiota by breaching the physical barrier. Epidemiologic studies in patients with scabies confirmed secondary bacterial infections by two clinically important pathogens *S. aureus* and *Streptococcus pyogenes*.³⁶

Lately, there has been a growing awareness of fungi and their interaction with the skin barrier. When the chemical composition (e.g., sweat, pH) of the host epidermis is disturbed, *Malassezia spp.* acquire pathogenicity and liberate an array of bioactive indoles, lipases, and phospholipases.³⁷ These molecules further modify the function of the skin barrier.

Epithelial barrier disruption opens the door

into a vicious itch–scratch cycle.^{38,39} Upon damage or stress, keratinocytes and skin microbiota emit cytokines, AMPs, and proteases that activate immunocytes and nerves.^{38,40,41} Protease-activated receptors (PARs), which are cleaved by serine proteases, manifest on different cell types, including sensory neurons and mediate itch.^{42–45} β -defensin, an AMP released from epithelial cells, has the ability to stimulate IL-31 production by mast cells.⁴⁶ IL-31, initially discovered in 2004, is the first cytokine that is known to facilitate itch by directly operating on sensory neurons (Figure 1).⁴⁷

The immune system. Skin is flushed with a wide scope of cells of the innate and adaptive immune system. The skin microbiota keeps immune homeostasis¹⁹ by modulating the expression of diverse innate factors, including AMPs, interleukin 1a (IL-1a),⁴⁸ and complement.⁴⁹ Symbionts calibrate inflammation.^{50,51} *S. epidermidis* suppresses inflammation by inducing IL-10, an anti-inflammatory cytokine, from antigen-presenting cells (APCs).⁵² The Toll-like-receptor (TLR)–2-facilitated recognition of lipoteichoic acid (LTA) from *S. epidermidis* inhibits TLR-3-driven inflammatory cytokine production in cultured keratinocytes (Table 1). This also reduces inflammation in wounds, which, when uncontrolled, is damaging to the host.⁵² Finally, *S. epidermidis* can finely tune the response of resident T cells and promote selective immunity against skin pathogens.⁵⁷

Alteration in the normal makeup of the skin microbiota can induce inflammation. Moreover, the constitution of the cutaneous microbiota can shift dramatically in the course of inflammation.¹⁴ For example, AD flares have been associated with an expansion of staphylococcal species, which can lead to an overall decrease in microbial diversity.¹⁴ The resulting bacterial and viral infection can cause itch. One possible mechanism of itch from *S.*

TABLE 1. Interaction between the skin microbiota and the Toll-like receptors (TLRs).

BACTERIA	INTERACTIONS WITH TLRs
<i>S. epidermidis</i>	Adjusts TLR3-dependent inflammation by introducing a TLR2-mediated crosstalk to subdue inflammation. ⁵²
<i>S. aureus</i>	Elicits keratinocytes to display AMPs through a TLR2-dependent mechanism. ⁵⁰ Induction of h β D3 gene expression is TLR2-dependent. ⁵³ Lipoteichoic acid and bacterial lipoproteins act as TLR2/2 or TLR2/6 agonists. ^{54,55}
<i>P. acnes</i>	Colonizes sebaceous glands and stimulates keratinocytes to release inflammatory cytokines via TLR2 activation. ⁵⁶

aureus infection is mast cell-mediated pruriceptor stimulation. Nunez et al⁵⁸ discovered that *S. aureus* releases delta-toxin, an amphipathic peptide that stimulates chemical release from mast cells and mediates skin pathology in AD. Serine protease from *S. aureus* is also involved in type-2 inflammation and itch.^{16,59}

Varicella zoster virus (VZV) causes pruritus in chickenpox by mast-cell-derived histamine.⁶⁰ Keratinocytes first detect pathogens and initiate an immune response.⁶¹ Keratinocytes identify an array of microbial ligands via Toll-like receptors (TLRs) exhibited on their surface.^{62–64} In response to stimulation, keratinocytes produce alarmins or epithelial cell-derived cytokines (i.e., IL-33, thymic stromal lymphopoietin [TSLP]),⁶⁵ which potentiate innate and adaptive immunity.⁶¹ TSLP also acts upon a subdivision of TRPA1 sensory neurons to spark itch.⁶⁵

Mast cells are also an essential element of innate immunity. Mast cells recognize pathogens via pathogen-associated molecular pattern (PAMP) receptors (e.g., TLR) on their surface.⁶⁶ Once they detect pathogens, inflammatory mediators are released to attract other immune cells.^{67,68} Downstream of IL-33 and TSLP, mast cells, neutrophils, basophils, eosinophils, T helper-2 (T_H2) cells, and macrophages generate cytokines (IL-4, IL-13, IL-31), histamine, proteases, serotonin (5-HT), lipids, S100 proteins, prostaglandin E₂ (PG-E₂), leukotriene B₄ (LT-B₄), and growth factors.^{69–71} Recognizing these pro-inflammatory molecules via TRPV1 and TRPA1 channels leads to potential propagation across the afferent itch pathway.⁷²

T_H2 immunity is dominant in scabies and is complemented by a heavy inflow of IL-31(+) M₂ macrophages.⁷³ Proteases from scabies mite stir epidermal keratinocytes to express TSLP. TSLP activates T_H2 cells and induces M₂ macrophages to produce IL-31, causing severe itch.⁷⁴ The antigens of *S. aureus* have also been reported to induce IL-31 in individuals with AD.⁷⁵

The sensory nerve. The skin is one of the human body's first lines of defense against microbial threats. Though the immune system is an essential component of cutaneous immunity, it is evident that the sensory nervous system also plays an important role in host defense. By evoking the sensation of itch, the host can immediately sense danger and rapidly initiate a protective behavioral response.⁶⁹

A network of high- and low-threshold sensory nerves innervates the skin and is frequently

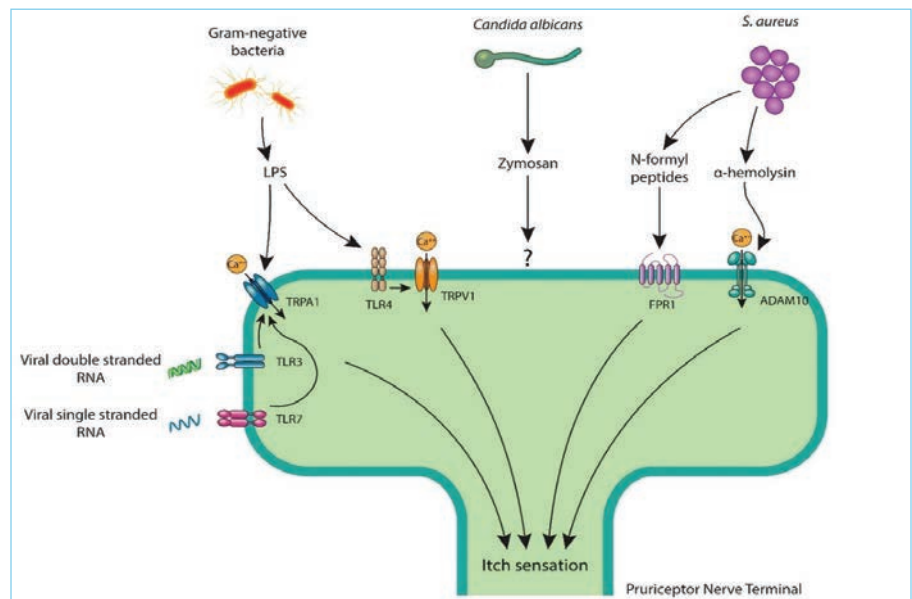


FIGURE 2. Pruriceptor neurons recognize skin pathogens and their molecular ligands by various mechanisms to facilitate itch. LPS, a key cell wall component of Gram-negative bacteria attaches to neuronal TLR4 and primes TRPV1 ion channel or opens the TRPA1 ion channel. *S. aureus* triggers itch with bacterial N-formyl peptides that bind to FPR1 or via α -hemolysin, which couples with ADAM10. *C. albicans* stimulates pruriceptors with its cell wall component zymosan. Viral double-strand RNA and single-strand RNA bind to TLR3 and TLR7, respectively, which are believed to sensitize the TRPA1 ion channel. ADAM10: a disintegrin and metalloproteinase domain-containing protein 10; FPR1: formyl peptide receptor 1; LPS: lipopolysaccharide; RNA: ribonucleic acid; TLR: Toll-like receptor; TRPA1: transient receptor potential ankyrin 1; TRPV1: transient receptor potential vanilloid 1.

exposed to bacterial pathogens (Figure 2). Pruriceptor neurons express cytokine receptors and G protein-coupled receptors that recognize immune mediators.⁷⁶ While it is understood that microbial inflammation propagates itch, how the skin microbiota directly triggers sensory nerves is a new area of inquiry. The latest studies suggest that sensory neurons (e.g., immune cells) are able to detect microbiota.^{13,69,76,77} Ji et al⁷⁸ reported TLR7 on pruriceptors and noted synthetic TLR7 ligands (i.e., imiquimod) causing itch behavior in mice.⁷⁸ TLR3 is also displayed by pruriceptors, where PolyI:C, a TLR3 ligand, stimulates neuronal activity and itch.⁷⁹ Viral single-stranded RNA and doublestranded RNA are known pathogen-derived ligands for TLR7 and TLR3, respectively, and there is a possibility that these viral ligands cause itch by directly interacting with pruriceptor neurons.⁷⁶

Lipopolysaccharide (LPS), an important component of the Gram-negative bacteria outer membrane binds to TLR4.⁸⁰ Although LPS has only been reported with pain,⁸¹ it can also modulate itch since TLR4 promotes histamine-mediated itch.⁸² Interestingly, LPS has also been found to stimulate sensory neurons in an

TLR4-independent manner, via the activation of TRPA1.^{83,84}

Pruriceptor neurons express cytokine receptors and G protein-coupled receptors that recognize immune mediators.⁷⁶ While we understand that microbial inflammation propagates itch, how the skin microbiota directly triggers sensory nerves is a new area of inquiry.

The latest studies suggest that sensory neurons, like immune cells, are able to detect microbiota.^{13,69,76,77} Ji et al⁷⁸ noted Toll-like receptor 7 (TLR7) on pruriceptors and synthetic TLR7 ligands (e.g., imiquimod) causing itch behavior in mice.⁷⁸ TLR3 is also displayed by pruriceptors, where PolyI:C, a TLR3 ligand, stimulates neuronal activity and itch.⁷⁹ Viral single-stranded ribonucleic acid (RNA) and double-stranded RNA are known pathogen-derived ligands for TLR7 and TLR3, respectively, and there is a possibility that these viral ligands cause itch by directly interacting with pruriceptor neurons.⁷⁶

Lipopolysaccharide (LPS), an important component of the Gram-negative bacteria outer membrane, binds to TLR4.⁸⁰ Although LPS has only been reported with pain,⁸¹ it can also modulate itch due to TLR4's promotion of

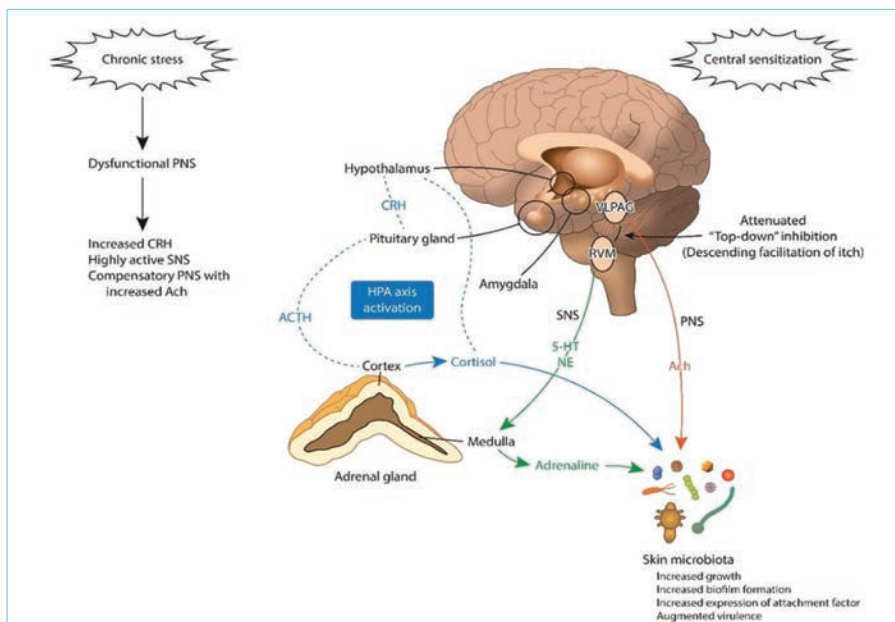


FIGURE 3. Brain to microbiota communication under chronic stress. The HPA axis is activated under chronic stress. The final product of the HPA axis, cortisol, directly activates skin microbes. Cortisol activates the amygdala, promoting central sensitization to itch. The amygdala also promotes CRH signaling to the brain stem (PAG), altering the “descending itch modulatory system”. Prolonged exposure to cortisol, NE, and ACTH is associated with increased growth and biofilm genesis and augmented virulence of the skin microbiota. Ach: acetylcholine; ACTH: adrenocorticotropic hormone; CRH: corticotropin-releasing-hormone; HPA: hypothalamic–pituitary–adrenal axis; 5-HT: serotonin; NE: norepinephrine; PNS: parasympathetic nervous system; RVM: rostral ventromedial medulla; SNS: sympathetic nervous system; VLPAG: ventrolateral periaqueductal grey matter.

histamine-mediated itch.⁸² Interestingly, LPS has also been found to stimulate sensory neurons in an TLR4-independent manner, via the activation of TRPA1.^{83,84}

Besides TLR ligands, sensory neurons detect pathogens through various molecular means. Specifically, zymosan from *Candida albicans*,⁸⁵ N-formylated peptides and α -hemolysin from *S. aureus*,⁸⁶ and streptolysin S from *S. pyogenes*⁸⁷ were shown to mediate pain through direct neuronal stimulation. It remains to be discovered whether pruriceptors similarly detect these pathogens to elicit itch.

Itch is bothersome in patients with cholestatic liver disease.⁸⁸ Recently, alteration of the skin microbiota was identified in patients with cirrhosis where specified microbial taxa correlated with itch severity and serum autotaxin (ATX) level.⁸⁹ Lysophosphatidic acid (LPA), a powerful neuronal activator, and ATX (ectonucleotide pyrophosphatase/ phosphodiesterase 2), the enzyme that creates LPA, are pruritogens in cholestasis.^{90,91} It has been suggested that LPA directly activates TRPV1 on peripheral sensory neurons to mediate itch.⁹²

Neuroimmune conversation is bidirectional in

itch. Sensory neurons are sensitized by immune cell-made cytokines, such as TNF- α and IL-1 β ; chemicals, such as histamine; and lipid mediators, such as prostaglandins; which phosphorylate ion channels and lower the bar of action potential firing. Neurons, in turn, secrete neuropeptides (e.g., calcitonin gene-related peptide, substance P) that modulate immune cell function^{93,94} and microbial virulence,^{95–97} causing itch.⁹⁸ Because neurons will respond within milliseconds to danger, the sensory nervous system is likely the

body's first detector of pathogen invasion and the prime orchestrator of itch.⁷⁶

THE CENTRAL MECHANISM LINKING THE SKIN MICROBIOTA AND ITCH

Microbial endocrinology. Microbial endocrinology is a combination of two distinct areas of study—microbiology and neurobiology—and is based on the shared presence of neurochemicals in the host and the microbiota.⁶⁶ The scope of neurochemicals and the variety of microbiota in which they have been discovered is expansive,⁹⁹ including acetylcholine,^{100,101} histamine,^{102,103} serotonin,¹⁰⁴ catecholamines,^{105,106} and agmatine,^{107,108} which are essential elements of an animal's nervous system. Others, such as corticotropin,¹⁰⁹ somatostatin,¹¹⁰ and progesterone,¹¹¹ have biological action in mammalian cells. The ability of the microbiota to not only respond to but also create the very same neurochemicals found in mammalian systems indicates that host interplay with its microbiota is more interactive than was previously thought. Hence, microbial endocrinology could potentially be applied beyond the study of infectious disease to other conditions, such as neurological disease, through the microbiota–skin–brain axis. Microbiota has multiple transmission pathways to access the brain, including the neural signals carried by the afferent neurons, endocrine messages transmitted by neurochemicals, and the immune response messages transferred by cytokines.^{112,113} Supporting cutaneous microbiota improves the skin's barrier functioning and local immune system and assists in its communication with other organ systems, including the brain (microbiota–skin–brain axis).¹¹⁴

Stress. Stress is a complex, dynamic event

TABLE 2. Effects of stress mediators on the skin microbiota.

BACTERIA	EFFECTS OF STRESS MEDIATORS
<i>Staphylococcus epidermidis</i>	Glucocorticoids decrease the effects of super antigen activated T cells and inhibit staphylococcal exotoxin-induced T cell proliferation, cytokine secretion. ¹³⁷ Catecholamines induce biofilm growth. ¹³⁰
<i>Propionibacterium acnes</i>	Cortisol and steroids significantly exacerbate inflammation associated with <i>P. acnes</i> via TLR2 stimulation. ^{138,139}
<i>Pseudomonas aeruginosa</i>	Norepinephrine increases expression of the attachment factor PA-1 of <i>P. aeruginosa</i> and increase biofilm formation. ^{135,138}
<i>Staphylococcus aureus</i>	Acetylcholine augments susceptibility to infection by <i>S. aureus</i> . ¹²⁴ Norepinephrine increases <i>S. aureus</i> ' ability to remove iron from host and therefore facilitates the bacteria to form biofilms. ^{138,140}
Group A <i>Streptococcus</i>	Cortisol alters vulnerability to Group A <i>Streptococcus pyogenes</i> skin infection. ¹⁴¹ Acetylcholine augments susceptibility to infection by Group A <i>Streptococcus</i> . ¹²⁴ Catecholamines raise Staphylococcal growth by 5-log orders. ^{130–132} Catecholamines enhance Group A <i>Streptococcus</i> growth likely by increasing iron availability. ^{138,142}
<i>Candida</i>	Estrogen enhances <i>Candida</i> infectivity, switching yeast form to an invasive hyphae. ¹⁴³

that alters the body's homeostasis and elicits a response in the host. Stress can aggravate itch,^{115–117} which indicates that the brain is engaged in the final common stage of itch processing.^{118,119} The stress response by the central nervous system (CNS) can alter the microbiota via the release of neurochemicals.^{120,121} Glucocorticoids, essential components of the stress response, repress AMP release/localization in the epidermis, weaken the barrier, and raise host susceptibility to infection.^{122–124} Chronic stress is associated with an aberrant parasympathetic tone (Figure 3).^{125,126} Cholinergic signaling from physiologic stress¹²⁵ can negatively impact the skin barrier and immunity.^{127,128} Cathelicidin and β -defensins, AMPs important for innate immunity, are cut down after $\alpha 7$ nAChR stimulation,^{128,129} leading to bacterial dissemination (Figure 3).^{127,128} Skin microbiota, especially the coagulase-negative staphylococci, are sensitive to catecholamines. Norepinephrine (NE), epinephrine, dopamine, and their structurally related inotropes (dobutamine and isoprenaline) raise staphylococcal growth by 5-log orders or more.^{130–132} Catecholamines also strengthen bacterial attachment to host tissues and increase bacteria virulence.^{130,133,134} Catecholamines stimulate the biofilm formation of *Pseudomonas aeruginosa* (*P. aeruginosa*) and *Escherichia coli* (*E. coli*).^{124,135} Within a polymicrobial biofilm, *P. aeruginosa* enhances USA300 MRSA virulence.¹³⁶ Substance P is released in sweat during stress and increases the virulence of Gram-positive skin bacteria, namely *S. aureus* and *S. epidermidis*.^{95,96} Thus, the effect of stress on the skin microbiota might be twofold: dampening the host defense to infection and causing changes to the microenvironment that make it more ideal for pathogens.¹²⁴ The resultant dysbiosis can exacerbate itch (i.e., “stress aggravated itch”) (Table 2).

The amygdala. Itch encompasses sensory-discriminative and affective-motivational aspects and undergoes extensive processing in the higher brain centers.^{119,125} The amygdala is involved in pain, especially in the emotional-affective aspects of pain perception.¹⁴⁴ The central nucleus of the amygdala (CeA) is commonly called the “nociceptive amygdala”¹⁴⁵ and receives peripheral pain signals via the parabrachial nucleus.¹⁴⁶ The role of amygdala in itch has also been shown in animal studies.¹⁴⁷ A recent study noted that scratching was suppressed after blocking itch-mediating spinal neurons connected to the spinoparabrachial pathway.¹⁴⁸ Additionally, an

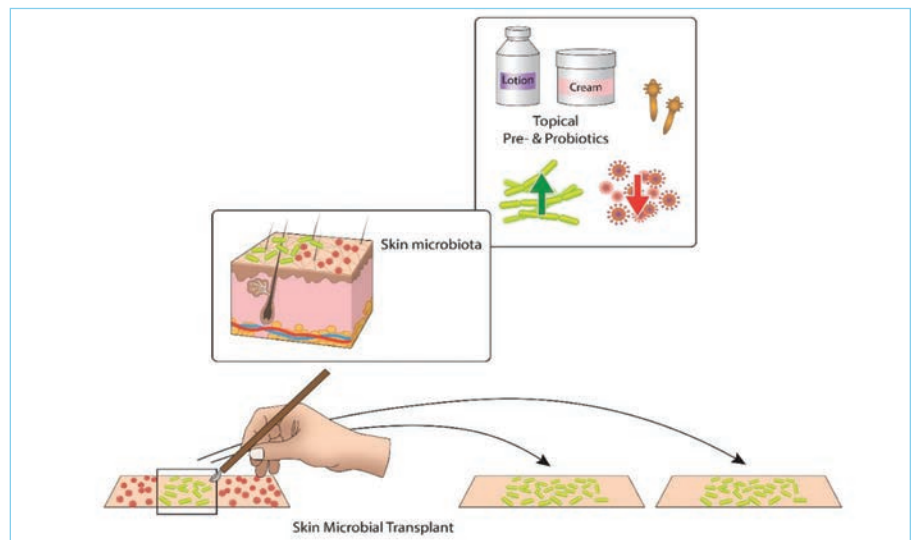


FIGURE 4. Two main approaches of controlling the human skin microbiota for the itch control. Topical pre- and probiotics target to increase the number of advantageous bacteria (green) and reduce pathogens (red). Skin microbial transplant is a new approach that transfers beneficial microbiota from healthy skin to itchy and dysbiotic skin.¹⁶⁷

animal functional MRI (fMRI) study demonstrated amygdala activation during itch stimuli.¹⁴⁹ The findings suggest that itch signals are delivered by both the spinothalamic pathway and the spinoparabrachial-CeA path. Injection of muscimol (γ -aminobutyric acid agonist) to the amygdala appeared to minimize the scratching elicited by the injection of serotonin to the cheek, suggesting a modulatory role of the amygdala in itch processing.¹⁵⁰ Chronic stress brings functional and configurational changes in the amygdala (central sensitization) (Figure 3).¹⁵¹ This change might influence itch processing in the brain, which might explain why stress can worsen itch in individuals with chronic itch.^{152,153} Studies suggest that the amygdala itself is susceptible to microbial influences.¹⁵⁴ Most convincingly, data from germ-free (GF) mice showed hyperactivity in the amygdala transcriptome in the absence of microbiota.^{155,156} This hyperactive state is in line with the altered pain sensitivity¹⁵⁷ and stress response in GF mice.^{158,159}

Currently, it is not clear how microbial signals navigate through the skin–brain axis to reach the amygdala specifically; however, there are some strong candidate paths, including the blood stream (circulation) and the spinal cord.^{112,154,160}

CONCLUSION AND FUTURE PERSPECTIVES

Increased recognition and understanding of the presence and functionality of the microbiota has changed what we know about the human body. Cutaneous microbiota appear to have a

diverse and far-reaching influence on human physiology by calling upon the host nervous system. Bacteria produce metabolites, toxins, and structural components that are recognized by peripheral and central neurons via matching receptors. Microbiota also appear to indirectly affect neural function by causing endocrine (e.g., keratinocytes) and immune cells to transmit signals (e.g., cytokines, proteases). Itch is a prototypic sensory neural function, and microbiota appear to propel the itch–scratch cycle.

Some descriptive studies have differentiated the microbiota found in itchy skin versus those of healthy skin. While dysbiosis is found in various pathologies, their presence raises a “chicken-or-the-egg” type question in that it remains unclear if dysbiosis leads to disease or the underlying disease results in microbial imbalance. To differentiate cause and effect, a deeper and more mechanistic (functional) understanding of the skin microbiota’s role in itch is required. Increased understanding will help us find microbiological markers in itchy conditions and develop more effective therapeutics that utilize host–microbiota relationship. The gut and skin are uniquely related in function, and numerous studies link gut microbiota to skin homeostasis (skin–gut axis or skin–gut–brain axis).^{35,161–164} Commonalities have also been found between itch transition in the skin and neural signaling in the lower intestinal tract, which raises the question of whether intestinal microbiota also

play a role in itching.^{165,166}

The interplay between the skin microbiota and itch is an emerging area of research with many potential areas of focus for therapeutic development: 1) Whole microbiota transplant, a process that offers microbiota from healthy donors to patients with significant skin dysbiosis, such as AD; 2) Topical probiotics to increase or introduce advantageous microbiota in patients, especially at a critical age for immune and limbic brain wiring; 3) Topical prebiotics to stimulate beneficial skin microbiota, or biomass or dead extracts of nonpathogenic bacteria to antagonize substance P; and 4) Studies on host–microbiota interplay that analyze microbial metabolites, re-imposing commensal microbial activity by offering signaling molecules (Figure 4).

In future years, we might see topical microbiota modulator cosmetics/transdermal drugs emerge that improve our health as well as our appearance.

AUTHOR CONTRIBUTIONS

All authors made substantial contributions to the concept and design of the work and approved the submitted version (Conceptualization, H.S.K., and G.Y.; Writing–Original Draft Preparation, Review and Editing, H.S.K., and G.Y.; Funding Acquisition, H.S.K.). All authors have read and agreed to the published version of the manuscript.

CREATIVE COMMONS

This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated.

REFERENCES

- Sender R, Fuchs S, Milo R. Are we really vastly outnumbered? Revisiting the ratio of bacterial to host cells in humans. *Cell*. 2016;164:337–340.
- Bianconi E, Piovesan A, Facchin F, et al. An estimation of the number of cells in the human body. *Ann Hum Biol*. 2013;40:463–471.
- Yang NJ, Chiu IM. Bacterial signaling to the nervous system through toxins and metabolites. *J Mol Biol*. 2017;429:587–605.
- Chen YE, Fischbach MA, Belkaid Y. Skin microbiota–host interactions. *Nature*. 2018;553:427–436.
- Gallo RL. Human skin is the largest epithelial surface for interaction with microbes. *J Invest Dermatol*. 2017;137:1213–1214.
- Fyhrquist N, Salava A, Auvinen P, et al. Skin biomes. *Curr Allergy Asthma Rep*. 2016;16:40.
- Sender R, Fuchs S, Milo R. Revised estimates for the number of human and bacteria cells in the body. *PLoS Biol*. 2016;14:e1002533.
- Leyden JJ, McGinley KJ, Nordstrom KM, et al. Skin microflora. *J Invest Dermatol*. 1987;88:655–725.
- Wang WM, Jin HZ. Skin microbiome: an actor in the pathogenesis of psoriasis. *Chin Med J (Engl)*. 2018;131:95–98.
- Trivedi B. Microbiome: the surface brigade. *Nature*. 2012;492:560–561.
- Gontcharova V, Youn E, Sun Y, et al. A comparison of bacterial composition in diabetic ulcers and contralateral intact skin. *Open Microbiol J*. 2010;4:8–19.
- Johnson TR, Gomez BI, McIntyre MK, et al. The cutaneous microbiome and wounds: new molecular targets to promote wound healing. *Int J Mol Sci*. 2018;19:2699.
- Chiu IM. Infection, pain, and itch. *Neurosci Bull*. 2018;34:109–119.
- Kong HH, Oh J, Deming C, et al. Temporal shifts in the skin microbiome associated with disease flares and treatment in children with atopic dermatitis. *Genome Res*. 2012;22:850–859.
- Blicharz L, Usarek P, Mlynarczyk G, et al. Is itch intensity in atopic dermatitis associated with skin colonization by *Staphylococcus aureus*? *Indian J Dermatol*. 2020;65:17–21.
- Allen HB, Vaze ND, Choi C, et al. The presence and impact of biofilm-producing *Staphylococci* in atopic dermatitis. *JAMA Dermatol*. 2014;150:260–265.
- Azimi E, Xia J, Lerner EA. Peripheral mechanisms of itch. *Curr Probl Dermatol*. 2016;50:18–23.
- Baldwin HE, Bhatia ND, Friedman A, et al. The role of cutaneous microbiota harmony in maintaining a functional skin barrier. *J Drugs Dermatol*. 2017;16:12–18.
- Sanford JA, Gallo RL. Functions of the skin microbiota in health and disease. *Semin Immunol*. 2013;25:370–377.
- Nakatsuji T, Chiang HI, Jiang SB, et al. The microbiome extends to subepidermal compartments of normal skin. *Nat Commun*. 2013;4:1431.
- Proksch E. pH in nature, humans and skin. *J Dermatol*. 2018;45:1044–1052.
- Boer M, Duchnik E, Maleszka R, et al. Structural and biophysical characteristics of human skin in maintaining proper epidermal barrier function. *Postepy Dermatol Alergol*. 2016;33:1–5.
- Niyonsaba F, Someya A, Hirata M, et al. Evaluation of the effects of peptide antibiotics human beta-defensins-1/-2 and LL-37 on histamine release and prostaglandin D(2) production from mast cells. *Eur J Immunol*. 2001;31:1066–1075.
- Capone KA, Dowd SE, Stamatias GN, et al. Diversity of the human skin microbiome early in life. *J Invest Dermatol*. 2011;131:2026–2032.
- Otto M. *Staphylococcus epidermidis*—the ‘accidental’ pathogen. *Nat Rev Microbiol*. 2009;7:555–567.
- Mah TF, O’Toole GA. Mechanisms of biofilm resistance to antimicrobial agents. *Trends Microbiol*. 2001;9:34–39.
- Iwase T, Uehara Y, Shinji H, et al. *Staphylococcus epidermidis* Espinhibits *Staphylococcus aureus* biofilm formation and nasal colonization. *Nature*. 2010;465:346–349.
- Shu M, Wang Y, Yu J, et al. Fermentation of

- Propionibacterium acnes*, a commensal bacterium in the human skin microbiome, as skin probiotics against methicillin-resistant *Staphylococcus aureus*. *PLoS ONE*. 2013;8:e55380.
- Grice EA, Segre JA. The skin microbiome. *Nat Rev Microbiol*. 2011;9:244–253.
- Korting HC, Hubner K, Greiner K, et al. Differences in the skin surface pH and bacterial microflora due to the long-term application of synthetic detergent preparations of pH 5.5 and pH 7.0. Results of a crossover trial in healthy volunteers. *Acta Derm Venereol*. 1990;70:429–431.
- Ali SM, Yosipovitch G. Skin pH: From basic science to basic skin care. *Acta Derm Venereol*. 2013;93:261–267.
- Van Smeden J, Bouwstra JA. Stratum corneum lipids: their role for the skin barrier function in healthy subjects and atopic dermatitis patients. *Curr Probl Dermatol*. 2016;49:8–26.
- Baker BS. The role of microorganisms in atopic dermatitis. *Clin Exp Immunol*. 2006;144:1–9.
- Ohnishi Y, Okino N, Ito M, et al. Ceramidase activity in bacterial skin flora as a possible cause of ceramide deficiency in atopic dermatitis. *Clin Diagn Lab Immunol*. 1999;6:101–104.
- Kim JE, Kim HS. Microbiome of the skin and gut in atopic dermatitis (AD): understanding the pathophysiology and finding novel management strategies. *J Clin Med*. 2019;8(4):444.
- Swe PM, Zakrzewski M, Kelly A, et al. Scabies mites alter the skin microbiome and promote growth of opportunistic pathogens in a porcine model. *PLoS Negl Trop Dis*. 2014;8:e2897.
- Xu J, Saunders CW, Hu P, et al. Dandruff-associated *Malassezia* genomes reveal convergent and divergent virulence traits shared with plant and human fungal pathogens. *Proc Natl Acad Sci USA*. 2007;104:18730–18735.
- Mack MR, Kim BS. The itch-scratch cycle: a neuroimmune perspective. *Trends Immunol*. 2018;39:980–991.
- Potenzieri C, Udem BJ. Basic mechanisms of itch. *Clin Exp Allergy*. 2012;42:8–19.
- Borgono CA, Michael IP, Komatsu N, et al. A potential role for multiple tissue kallikrein serine proteases in epidermal desquamation. *J Biol Chem*. 2007;282:3640–3652.
- Yosipovitch G, Misery L, Proksch E, et al. Skin barrier damage and itch: review of mechanisms, topical management and future directions. *Acta Derm Venereol*. 2019;99:1201–1209.
- Komatsu N, Saijoh K, Kuk C, et al. Human tissue kallikrein expression in the stratum corneum and serum of atopic dermatitis patients. *Exp Dermatol*. 2007;16:513–519.
- Steinhoff M, Neisius U, Ikoma A, et al. Proteinase-activated receptor-2 mediates itch: a novel pathway for pruritus in human skin. *J Neurosci*. 2003;23:6176–6180.
- Stefansson K, Brattsand M, Roosterman D, et al. Activation of proteinase-activated receptor-2 by human kallikrein-related peptidases. *J Invest Dermatol*. 2008;128:18–25.
- Sanders KM, Nattkemper LA, Rosen JD, et al. Non-histaminergic itch mediators elevated in the skin of a porcine model of scabies and of human scabies patients. *J Invest Dermatol*. 2019;139:971–973.
- Niyonsaba F, Ushio H, Hara M, et al. Antimicrobial peptides human beta-defensins and cathelicidin LL-37 induce the secretion of a pruritogenic cytokine IL-31 by human mast cells. *J Immunol*. 2010;184:3526–3534.
- Cevikbas F, Wang X, Akiyama T, et al. A sensory neuron-expressed IL-31 receptor mediates T helper cell-dependent itch: involvement of TRPV1 and TRPA1. *J Allergy Clin Immunol*. 2014;133:448–460.
- Naik S, Bouladoux N, Wilhelm C, et al. Compartmentalized control of skin immunity by resident commensals. *Science*. 2012;337:1115–1119.

49. Chehoud C, Rafail S, Tyldsley AS, et al. Complement modulates the cutaneous microbiome and inflammatory milieu. *Proc Natl Acad Sci USA*. 2013;110:15061–15066.
50. Lai Y, Cogen AL, Radek KA, et al. Activation of TLR2 by a small molecule produced by *Staphylococcus epidermidis* increases antimicrobial defense against bacterial skin infections. *J Invest Dermatol*. 2010;130:2211–2221.
51. Stacy A, Belkaid Y. Microbial guardians of skin health. *Science*. 2019;363:227–228.
52. Lai Y, Di Nardo A, Nakatsuji T, et al. Commensal bacteria regulate Toll-like receptor 3-dependent inflammation after skin injury. *Nat Med*. 2009;15:1377–1382.
53. Menzies BE, Kenoyer A. Signal transduction and nuclear responses in *Staphylococcus aureus*-induced expression of human beta-defensin 3 in skin keratinocytes. *Infect Immun*. 2006;74:6847–6854.
54. Hashimoto M, Tawaratsumida K, Kariya H, et al. Lipoprotein is a predominant Toll-like receptor 2 ligand in *Staphylococcus aureus* cell wall components. *Int Immunol*. 2006;18:355–362.
55. Bubeck-Wardenburg J, Williams WA, Missiakos D. Host defenses against *Staphylococcus aureus* infection require recognition of bacterial lipoproteins. *Proc Natl Acad Sci USA*. 2006;103:13831–13836.
56. Kim J, Ochoa, MT, Krutzyk SR, et al. Activation of Toll-like receptor 2 in acne triggers inflammatory cytokine responses. *J Immunol*. 2002;169:1535–1541.
57. Linehan JL, Harrison OJ, Han SJ, et al. Non-classical immunity controls microbiota impact on skin immunity and tissue repair. *Cell*. 2018;172:784–796.e18.
58. Nakamura Y, Oscherwitz J, Cease KB, et al. *Staphylococcus delta-toxin* induces allergic skin disease by activating mast cells. *Nature*. 2013;503:397–401.
59. Williams MR, Nakatsuji T, Gallo RL. *Staphylococcus aureus*: master manipulator of the skin. *Cell Host Microbe*. 2017;22:579–581.
60. Tebruegge M, Kuruvilla M, Margaron I. Does the use of calamine or antihistamine provide symptomatic relief from pruritus in children with varicella zoster infection? *Arch Dis Child*. 2006;91:1035–1036.
61. McKenzie RC, Sauder DN. Keratinocyte cytokines and growth factors. Functions in skin immunity and homeostasis. *Dermatol Clin*. 1990;8:649–661.
62. Kollich G, Kalali BN, Voelcker V, et al. Various members of the Toll-like receptor family contribute to the innate immune response of human epidermal keratinocytes. *Immunology*. 2005;114:531–541.
63. Lebre MC, Van der Aar AM, Van Baarsen, et al. Human keratinocytes express functional Toll-like receptor 3, 4, 5, and 9. *J Invest Dermatol*. 2007;127:331–341.
64. Olaru F, Jensen LE. Chemokine expression by human keratinocyte cell lines after activation of Toll-like receptors. *Exp Dermatol*. 2010;19:e314–e316.
65. Dainichi T, Kitoh A, Otsuka A, et al. The epithelial immune microenvironment (EIME) in atopic dermatitis and psoriasis. *Nat Immunol*. 2018;19:1286–1298.
66. Hofmann AM, Abraham SN. New roles for mast cells in modulating allergic reactions and immunity against pathogens. *Curr Opin Immunol*. 2009;21:679–686.
67. Igawa S, Di Nardo A. Skin microbiome and mast cells. *Transl Res*. 2017;184:68–76.
68. Leon A, Rosen JD, Hashimoto T, et al. Itching for an answer: a review of potential mechanisms of scalp itch in psoriasis. *Exp Dermatol*. 2019;28:1397–1404.
69. Trier AM, Mack MR, Kim BS. The neuroimmune axis in skin sensation, inflammation, and immunity. *J Immunol*. 2019;202:2829–2835.
70. Walsh CM, Hill RZ, Schwendinger-Schreck J, et al. Neutrophils promote CXCR3-dependent itch in the development of atopic dermatitis. *Elife*. 2019;8.
71. Hashimoto T, Rosen JD, Sanders KM, et al. Possible role of neutrophils in itch. *Itch*. 2018;3:e17.
72. Luo J, Feng J, Liu S, et al. Molecular and cellular mechanisms that initiate pain and itch. *Cell Mol Life Sci*. 2015;72:3201–3223.
73. Hashimoto T, Satoh T, Yokozeki H. Pruritus in ordinary scabies: IL-31 from macrophages induced by overexpression of thymic stromal lymphopoietin and perioest. *Allergy*. 2019;74:1727–1737.
74. Hashimoto T, Kursewicz CD, Fayne RA, et al. Mechanisms of itch in stasis dermatitis: significant role of IL-31 from macrophages. *J Invest Dermatol*. 2020;140:850–859.
75. Sonkoly E, Muller A, Lauerman AI, et al. IL-31: a new link between T cells and pruritus in atopic skin inflammation. *J Allergy Clin Immunol*. 2006;117: 411–417.
76. Baral P, Mills K, Pinho-Ribeiro FA, et al. Pain and itch: beneficial or harmful to antimicrobial defense? *Cell Host Microbe*. 2016;19:755–759.
77. Liu T, Gao YJ, Ji RR. Emerging role of Toll-like receptors in the control of pain and itch. *Neurosci Bull*. 2012;28: 131–144.
78. Liu T, Xu ZZ, Park CK, et al. Toll-like receptor 7 mediates pruritus. *Nat Neurosci*. 2010;13:1460–1462.
79. Liu T, Berta T, Xu ZZ, et al. TLR3 deficiency impairs spinal cord synaptic transmission, central sensitization, and pruritus in mice. *J Clin Invest*. 2012;122:2195–2207.
80. Diogenes A, Ferraz CC, Akopian AN, et al. LPS sensitizes TRPV1 via activation of TLR4 in trigeminal sensory neurons. *J Dent Res*. 2011;90:759–764.
81. Calil IL, Zarpelon AC, Guerrero AT, et al. Lipopolysaccharide induces inflammatory hyperalgesia triggering a TLR4/MyD88-dependent cytokine cascade in the mice paw. *PLoS ONE*. 2014;9:e90013.
82. Min H, Lee H, Lim H, et al. TLR4 enhances histamine-mediated pruritus by potentiating TRPV1 activity. *Mol Brain*. 2014;7:59.
83. Meseguer V, Alpizar YA, Luis E, et al. TRPA1 channels mediate acute neurogenic inflammation and pain produced by bacterial endotoxins. *Nat Commun*. 2014;5:3125.
84. Startek JB, Talavera K, Voets T, et al. Differential interactions of bacterial lipopolysaccharides with lipid membranes: implications for TRPA1-mediated chemosensation. *Sci Rep*. 2018;8:12010.
85. Kashem SW, Riedl MS, Yao, C, et al. Nociceptive sensory fibers drive interleukin-23 production from CD301b+ dermal dendritic cells and drive protective cutaneous immunity. *Immunity*. 2015;43:515–526.
86. Chiu IM, Heesters BA, Ghasemlou N, et al. Bacteria activate sensory neurons that modulate pain and inflammation. *Nature*. 2013;501:52–57.
87. Pinho-Ribeiro FA, Baddal B, Haarsma R, et al. Blocking neuronal signaling to immune cells treats Streptococcal invasive infection. *Cell*. 2018;173:1083–1097.e22.
88. Hashimoto T, Yosipovitch G. Itching as a systemic disease. *J Allergy Clin Immunol*. 2019;144:375–380.
89. Bajaj JS, Fagan A, Sikaroodi M, et al. Alterations in skin microbiomes of patients with cirrhosis. *Clin Gastroenterol Hepatol*. 2019;17:2581–2591.e15.
90. Kremer AE, Martens JJ, Kulik W, et al. Lysophosphatidic acid is a potential mediator of cholestatic pruritus. *Gastroenterology*. 2010;139:1008–1018.e1.
91. Beuers U, Kremer AE, Bolier R, et al. Pruritus in cholestasis: facts and fiction. *Hepatology*. 2014;60:399–407.
92. Nieto-Posadas A, Picazo-Juarez G, Llorente I, et al. Lysophosphatidic acid directly activates TRPV1 through a C-terminal binding site. *Nat Chem Biol*. 2011;8:78–85.
93. Roosterman D, Goerge T, Schneider SW, et al. Neuronal control of skin function: the skin as a neuroimmunoendocrine organ. *Physiol Rev*. 2006;86:1309–1379.
94. Pereira U, Boulais N, Lebonvallet N, et al. Development of an in vitro coculture of primary sensitive pig neurons and keratinocytes for the study of cutaneous neurogenic inflammation. *Exp Dermatol*. 2010;19:931–935. *J Clin Med*. 2020;9:1190.
95. Mijouin L, Hillion M, Ramdani Y, et al. Effects of a skin neuropeptide (substance P) on cutaneous microflora. *PLoS ONE*. 2013;8:e78773.
96. N'Diaye A, Gannesen A, Borrel V, et al. Substance P and calcitonin gene-related peptide: key regulators of cutaneous microbiota homeostasis. *Front Endocrinol (Lausanne)*. 2017;8:15.
97. N'Diaye A, Mijouin L, Hillion M, et al. Effect of substance P in *Staphylococcus aureus* and *Staphylococcus epidermidis* virulence: implication for skin homeostasis. *Front Microbiol*. 2016;7:506.
98. Raap U, Stander S, Metz M. Pathophysiology of itch and new treatments. *Curr Opin Allergy Clin Immunol*. 2011;11:420–427.
99. Lenard J. Mammalian hormones in microbial cells. *Trends Biochem Sci*. 1992;17:147–150.
100. Kawashima K, Misawa H, Moriwaki Y, et al. Ubiquitous expression of acetylcholine and its biological functions in life forms without nervous systems. *Life Sci*. 2007;80: 2206–2209.
101. Stephenson M, Rowatt E. The production of acetylcholine by a strain of *Lactobacillus plantarum*. *J Gen Microbiol*. 1947;1:279–298.
102. Masson F, Talon R, Montel MC. Histamine and tyramine production by bacteria from meat products. *Int J Food Microbiol*. 1996;32:199–207.
103. Thomas CM, Hong T, Van Pijkeren JP, et al. Histamine derived from probiotic *Lactobacillus reuteri* suppresses TNF via modulation of PKA and ERK signaling. *PLoS ONE*. 2012;7:e31951.
104. Hurley R, Leask BG, Ruthven CR, et al. Investigation of 5-hydroxytryptamine production by *Candida albicans* in vitro and in vivo. *Microbios*. 1971;4:133–143.
105. Asano Y, Hiramoto T, Nishino R, et al. Critical role of gut microbiota in the production of biologically active, free catecholamines in the gut lumen of mice. *Am J Physiol Gastrointest Liver Physiol*. 2012;303: G1288–G1295.
106. Tsavkelova EA, Botvinko IV, Kudrin VS, et al. Detection of neurotransmitter amines in microorganisms with the use of high-performance liquid chromatography. *Dokl Biochem*. 2000;372:115–117.
107. Raasch W, Regunathan S, Li G, et al. Agmatine, the bacterial amine, is widely distributed in mammalian tissues. *Life Sci*. 1995;56:2319–2330.
108. Arena ME, Manca de Nadra MC. Biogenic amine production by *Lactobacillus*. *J Appl Microbiol*. 2001;90:158–162.
109. Leroith D, Liotta AS, Roth J, et al. Corticotropin and beta-endorphin-like materials are native to unicellular organisms. *Proc Natl Acad Sci USA*. 1982;79: 2086–2090.
110. LeRoith D, Pickens W, Vinik AI, et al. *Bacillus subtilis* contains multiple forms of somatostatin-like material. *Biochem Biophys Res Commun*. 1985;127:713–719.
111. Schar G, Stover EP, Clemons KV, et al. Progesterone binding and inhibition of growth in *Trichophyton mentagrophytes*. *Infect Immun*. 1986;52:763–767.
112. Cryan JF, Dinan TG. Mind-altering microorganisms: the impact of the gut microbiota on brain and behaviour. *Nat Rev Neurosci*. 2012;13:701–712.
113. O'Mahony SM, Dinan TG, Cryan JF. The gut microbiota as a key regulator of visceral pain. *Pain*. 2017;158(Suppl 1): S19–S28.
114. Holzer P, Farzi A. Neuropeptides and the microbiota-gut-

- brain axis. *Adv Exp Med Biol.* 2014;817:195–219.
115. Yosipovitch G, Ansari N, Goon A, et al. Clinical characteristics of pruritus in chronic idiopathic urticaria. *Br J Dermatol.* 2002;147:32–36.
116. Yosipovitch G, Goon A, Wee J, et al. The prevalence and clinical characteristics of pruritus among patients with extensive psoriasis. *Br J Dermatol.* 2000;143:969–973.
117. Yosipovitch G, Goon AT, Wee J, et al. Itch characteristics in Chinese patients with atopic dermatitis using a new questionnaire for the assessment of pruritus. *Int J Dermatol.* 2002;41:212–216.
118. Golpanian RS, Kim HS, Yosipovitch G. Effects of stress on itch. *Clin Ther.* 2020.
119. Yosipovitch G, Mochizuki H. Neuroimaging of itch as a tool of assessment of chronic itch and its management. *Handb Exp Pharmacol.* 2015;226:57–70.
120. Galley JD, Nelson MC, Yu Z, et al. Exposure to a social stressor disrupts the community structure of the colonic mucosa-associated microbiota. *BMC Microbiol.* 2014;14:189.
121. Bailey MT. Influence of stressor-induced nervous system activation on the intestinal microbiota and the importance for immunomodulation. *Adv Exp Med Biol.* 2014;817:255–276.
122. Slominski A. A nervous breakdown in the skin: stress and the epidermal barrier. *J Clin Invest.* 2007;117:3166–3169.
123. Aberg KM, Radek KA, Choi EH, et al. Psychological stress downregulates epidermal antimicrobial peptide expression and increases severity of cutaneous infections in mice. *J Clin Invest.* 2007;117:3339–3349.
124. Radek KA. Antimicrobial anxiety: the impact of stress on antimicrobial immunity. *J Leukoc Biol.* 2010;88:263–277.
125. Kim HS, Yosipovitch G. An aberrant parasympathetic response: a new perspective linking chronic stress and itch. *Exp Dermatol.* 2013;22:239–244.
126. Tran BW, Papoiu AD, Russoniello CV, et al. Effect of itch, scratching and mental stress on autonomic nervous system function in atopic dermatitis. *Acta Derm Venereol.* 2010;90:354–361.
127. Curtis BJ, Radek KA. Cholinergic regulation of keratinocyte innate immunity and permeability barrier integrity: new perspectives in epidermal immunity and disease. *J Invest Dermatol.* 2012;132:28–42.
128. Radek KA, Elias PM, Taupenot L. Neuroendocrine nicotinic receptor activation increases susceptibility to bacterial infections by suppressing antimicrobial peptide production. *Cell Host Microbe.* 2010;7:277–289.
129. Curtis BJ, Plichta JK, Blatt H, et al. Nicotinic acetylcholine receptor stimulation impairs epidermal permeability barrier function and recovery and modulates cornified envelope proteins. *Life Sci.* 2012;91:1070–1076.
130. Lyte M, Freestone PP, Neal CP, et al. Stimulation of *Staphylococcus epidermidis* growth and biofilm formation by catecholamine inotropes. *Lancet.* 2003;361:130–135.
131. Freestone PP, Haigh RD, Williams PH, et al. Stimulation of bacterial growth by heat-stable, norepinephrine-induced autoinducers. *FEMS Microbiol Lett.* 1999;172:53–60.
132. Neal CP, Freestone PP, Maggs AF, et al. Catecholamine inotropes as growth factors for *Staphylococcus epidermidis* and other coagulase-negative staphylococci. *FEMS Microbiol Lett.* 2001;194:163–169.
133. Borrel V, Thomas P, Catovic C, et al. Acne and stress: impact of catecholamines on *Cutibacterium acnes*. *Front Med (Lausanne).* 2019;6:155.
134. Clarke SR, Mohamed R, Bian L, et al. The *Staphylococcus aureus* surface protein IsdA mediates resistance to innate defenses of human skin. *Cell Host Microbe.* 2007;1:199–212.
135. Freestone PP, Sandrini SM, Haigh RD, et al. Microbial endocrinology: how stress influences susceptibility to infection. *Trends Microbiol.* 2008;16:55–64.
136. Pastar I, Nusbaum AG, Gil J, et al. Interactions of methicillin resistant *Staphylococcus aureus* USA300 and *Pseudomonas aeruginosa* in polymicrobial wound infection. *PLoS ONE.* 2013;8:e56846.
137. Choi EH, Demerjian M, Crumrine D, et al. Glucocorticoid blockade reverses psychological stress-induced abnormalities in epidermal structure and function. *Am J Physiol Regul Integr Comp Physiol.* 2006;291:R1657–R1662.
138. Sandrini SM, Shergill R, Woodward J, et al. Elucidation of the mechanism by which catecholamine stress hormones liberate iron from the innate immune defense proteins transferrin and lactoferrin. *J Bacteriol.* 2010;192:587–594.
139. Shibata M, Katsuyama M, Onodera T, et al. Glucocorticoids enhance Toll-like receptor 2 expression in human keratinocytes stimulated with *Propionibacterium acnes* or proinflammatory cytokines. *J Invest Dermatol.* 2009;129:375–382.
140. Seth AK, Geringer MR, Nguyen KT, et al. Bacteriophage therapy for *Staphylococcus aureus* biofilm-infected wounds: a new approach to chronic wound care. *Plast Reconstr Surg.* 2013;131:225–234.
141. Rojas IG, Padgett DA, Sheridan JF, et al. Stress-induced susceptibility to bacterial infection during cutaneous wound healing. *Brain Behav Immun.* 2002;16:74–84.
142. Cogen AL, Nizet V, Gallo RL. Skin microbiota: a source of disease or defence? *Br J Dermatol.* 2008;158:442–455.
143. Sonnex C. Influence of ovarian hormones on urogenital infection. *Sex Transm Infect.* 1998;74:11–19.
144. Veinante P, Yalcin I, Barrot M. The amygdala between sensation and affect: a role in pain. *J Mol Psychiatry.* 2013;1:9.
145. Neugebauer V, Li W. Differential sensitization of amygdala neurons to afferent inputs in a model of arthritic pain. *J Neurophysiol.* 2003;89:716–727.
146. Neugebauer V, Li W, Bird GC, et al. The amygdala and persistent pain. *Neuroscientist.* 2004;10:221–234.
147. Sanders KM, Akiyama T. The vicious cycle of itch and anxiety. *Neurosci Biobehav Rev.* 2018;87:17–26.
148. Mu D, Deng J, Liu KF, et al. A central neural circuit for itch sensation. *Science.* 2017;357:695–699.
149. Jeong KY, Kang JH. Investigation of the pruritus-induced functional activity in the rat brain using manganese-enhanced MRI. *J Magn Reson Imaging.* 2015;42:709–716.
150. Davidson S, Zhang X, Khasabov SG, et al. Relief of itch by scratching: state-dependent inhibition of primate spinothalamic tract neurons. *Nat Neurosci.* 2009;12:544–546.
151. Roozendaal B, McEwen BS, Chattarji S. Stress, memory and the amygdala. *Nat Rev Neurosci.* 2009;10:423–433.
152. Pavlenko D, Akiyama T. Why does stress aggravate itch? A possible role of the amygdala. *Exp Dermatol.* 2019;28:1439–1441.
153. Mochizuki H, Hernandez LE, Yosipovitch G. What does brain imaging tell us about itch? *Itch.* 2019;4:e23.
154. Cowan CSM, Hoban AE, Ventura-Silva AP, et al. Gutsy moves: the amygdala as a critical node in microbiota to brain signaling. *Bioessays.* 2018;40.
155. Stilling RM, Ryan FJ, Hoban AE, et al. Microbes & neurodevelopment—absence of microbiota during early life increases activity-related transcriptional pathways in the amygdala. *Brain Behav Immun.* 2015;50:209–220.
156. Hoban AE, Stilling RM, Moloney GM, et al. Microbial regulation of microRNA expression in the amygdala and prefrontal cortex. *Microbiome.* 2017;5:102.
157. Luczynski P, Tramullas M, Viola M, et al. Microbiota regulates visceral pain in the mouse. *Elife.* 2017;6.
158. Sudo N, Chida Y, Aiba Y, et al. Postnatal microbial colonization programs the hypothalamic-pituitary-adrenal system for stress response in mice. *J Physiol.* 2004;558:263–275.
159. Clarke G, Grenham S, Scully P, et al. The microbiome-gut-brain axis during early life regulates the hippocampal serotonergic system in a sex-dependent manner. *Mol Psychiatry.* 2013;18:666–673.
160. Onalapo OJ, Onalapo AY, Olowe AO. The neurobehavioral implications of the brain and microbiota interaction. *Front Biosci (Landmark Ed.).* 2020;25:363–397.
161. O'Neill CA, Monteleone G, McLaughlin JT, et al. The gut-skin axis in health and disease: a paradigm with therapeutic implications. *Bioessays.* 2016;38:1167–1176.
162. Salem I, Ramser A, Isham N, et al. The gut microbiome as a major regulator of the gut-skin axis. *Front Microbiol.* 2018;9:1459.
163. Arck P, Handjiski B, Hagen E, et al. Is there a 'gut-brain-skin axis'? *Exp Dermatol.* 2010;19:401–405.
164. Lee YB, Byun EJ, Kim HS. Potential role of the microbiome in acne: a comprehensive review. *J Clin Med.* 2019;8:987.
165. Sanders KM, Nattkemper LA, Yosipovitch G. The gut-itch connection. *Exp Dermatol.* 2016;25:344–345.
166. Castro J, Harrington AM, Lieu T, et al. Activation of pruritogenic TGR5, MrgprA3, and MrgprC11 on colon-innervating afferents induces visceral hypersensitivity. *JCI Insight.* 2019;4.
167. Egert M, Simmering R, Riedel CU. The association of the skin microbiota with health, immunity, and disease. *Clin Pharmacol Ther.* 2017;102:62–69.
168. Dreno B, Aravitskaia E, Berardesca E, et al. Microbiome in healthy skin, update for dermatologists. *J Eur Acad Dermatol Venereol.* 2016;30:2038–2047.
169. Bastiaansen TFS, Cowan CSM, Claessens MJ, et al. Making sense of... the microbiome in psychiatry. *Int J Neuropsychopharmacol.* 2019;22:37–52.
170. Davani-Davari D, Negahdarpour M, Karimzadeh I, et al. Prebiotics: definition, types, sources, mechanisms, and clinical applications. *Foods.* 2019;8:92. **NPPA**